# SEASONAL VARIATIONS IN TRANS-2-HEXENAL AND LINOLENIC ACID IN HOMOGENATES OF THEA SINENSIS LEAVES

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Key Word Index—Thea sinensis; Theaceae; tea; seasonal variations; cis-3-hexenal; trans-2-hexenal; cis-3-hexenal; linolenic acid.

Abstract—The seasonal variations in the amounts of C<sub>6</sub>-volatile components (cis-3-hexenal, trans-2-hexenal, n-hexanal) and their precursors (linoleic and linolenic acid) in homogenates of Thea sinensis leaves were quantitatively analyzed throughout the year. Formation of trans-2-hexenal began in the middle of April and reached a maximum during July. Towards autumn the aldehyde gradually decreased and, in winter (December to March), was virtually absent. The levels of cis-3-hexenol remained constant during May-December. cis-3-Hexenal showed a similar variation pattern to that of trans-2-hexenal. The major fatty acids in the leaves were palmitic, palmitoleic, oleic, linoleic and linolenic acid, and occurred in non-ionic lipids and phospholipid fractions. The amounts of linoleic and linolenic acid did not show any marked variation except for a big peak in October.

### INTRODUCTION

We have been studying the biosynthetic pathway of cis-3-hexenol (leaf alcohol) and trans-2-hexenal (leaf aldehyde) via the intermediate, cis-3-hexenal, from linolenic acid in homogenates of Thea sinensis leaves [1-11]. Using linolenic [ $U^{-14}C$ ] and linoleic [ $U^{-14}C$ ] acid we recently found that cis-3-hexenal, trans-2-hexenal and n-hexanal, were generated by an enzyme bound to lamellae of chloroplasts in tea leaves, when the leaves were cut or mechanically ruptured in the presence of  $O_2$  [11].

To demonstrate the formation of the C<sub>6</sub>-volatile components from linoleic and linolenic acid in homogenates of tea leaves throughout the year, the seasonal amounts of C<sub>6</sub>-volatile compounds and unsaturated fatty acids were quantitatively analyzed in the period 1974–1976.

# RESULTS AND DISCUSSION

V ariations in the amounts of  $C_6$ -unsaturated aldehydes and the corresponding alcohols

Fresh tea leaves were blended in air, the homogenates steam-distilled and the distillates extracted with Et<sub>2</sub>O; the crude essential oil thus obtained was quantitatively analyzed by GLC. The generation of cis-3-hexenal was followed by headspace vapour analysis according to the methods previously reported, as this aldehyde is easily isomerized to trans-2-hexenal in homogenates [1, 4]. The most prominent seasonal variations in the amounts of C<sub>6</sub>-aldehydes took place in the young leaves; the amount of trans-2-hexenal formed decreased from autumn to winter, after reaching a maximum in June-July (Fig. 1) During winter, the amount of trans-2-hexenal formed was extremely small and, in particular, was not appreciable from the end of December to the middle of April.

The seasonal variation pattern in the generation of cis-3-hexenal was similar to that of trans-2-hexenal (Fig. 2) and the amount of n-hexanal was smaller than that of cis-3-hexanal throughout the year. On the other hand, the amount of cis-3-hexenol formed was less than that of trans-2-hexenal throughout the year and showed only minor changes during May-December (Fig. 1).

Variation in the amounts of linoleic and linolenic acid

The results of the quantitative analysis of fatty acids constituting the non-ionic and phospholipid fractions in tea leaves are summarized in Table 1. The major fatty acids in the lipids were palmitic, palmitoleic, oleic, linoleic and linolenic acid, and they were not found in the free fatty acid fraction [12]. Of these acids linolenic acid was by far the most abundant. Linoleic and linolenic acids were more plentiful in the non-ionic fraction than in the phospholipid fraction, and unlike the C<sub>6</sub>-aldehydes did not show marked seasonal changes. The concentration of these acids was higher in the non-ionic lipid than in the phospholipid fraction throughout the year

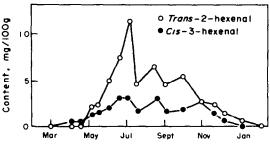


Fig. 1. Seasonal variation of volatile components in homogenates of fresh tea leaves.

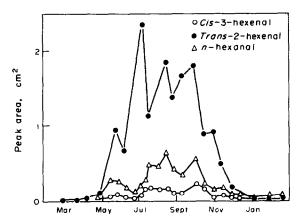


Fig. 2. Seasonal variation of volatile components in homogenates of fresh tea leaves.

(Fig. 3). The levels of these fatty acids in both the nonionic and phospholipid fractions notably decreased, corresponding to production of the flavour components on maceration or blending in the presence of  $O_2$ . The decrease in linolenic acid was large in the early summer and small in winter (Table 2). These variations in the decrease in unsaturated fatty acids during blending corresponded to the seasonal variations in the generation of  $C_6$ -unsaturated aldehydes (Figs. 1 and 2).

The C<sub>6</sub>-aldehyde forming activities reached a maximum in July-August and were very slight or absent during January-March, paralleling the seasonal variations of C<sub>6</sub>-volatile components in the leaves [13].

## **EXPERIMENTAL**

Fresh leaves of Thea sinensis var. Yahukita grown in Yamaguchi University were harvested from May 1974 to March

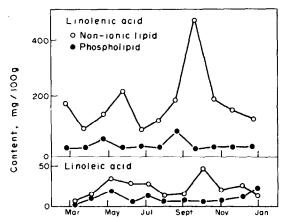


Fig. 3. Seasonal variation of linolenic and linoleic acids in non-ionic and phospholipid fractions in homogenates of tea leaves.

1976. Linolenic and linoleic acids were high-purity grade of above 98%. Authentic specimens of *n*-hexanal, *cis*-3-hexenal and *trans*-2-hexenal were synthesized through unequivocal routes [6].

Preparation of essential oil. Fresh leaves (150 g) were blended with H<sub>2</sub>O (500 ml) for 3 min in air, and the homogenate was steam-distilled to give 500 ml of distillate. Distillate was extracted with 3 × 50 ml of Et<sub>2</sub>O after saturation with NaCl and the combined extracts dried. Evaporation of the solvent afforded 20 mg of crude essential oil. A portion of the oil was quantitatively analyzed by FID GLC using a 3 mm × 3 m stainless steel column packed with 20% PEG-20 M on Celite 545; column temp, 120°C; injection temp, 150°; flow rate N<sub>2</sub>, 80 ml/min.

Preparation of vapour sample. Fresh leaves (5 g) were blended for 2 min with H<sub>2</sub>O (80 ml) in air, and the homogenate was transferred immediately to a rubber-stoppered conical flask (50 ml) and incubated at 25° for 5 min, then at

Table 1. Composition of fatty acids in lipid fractions from tea leaves

| Fatty acid  | Free fatty<br>acid | Fraction<br>Non-ionic<br>lipid | Phospholipid | Total |
|-------------|--------------------|--------------------------------|--------------|-------|
| Palmitic    | trace              | 38.7*                          | 14.5         | 53.2  |
| Palmitoleic | trace              | 5.8                            | trace        | 5.8   |
| Oleic       | trace              | 5.0                            | 3.4          | 8.4   |
| Linoleic    | trace              | 18.5                           | 13.8         | 32.3  |
| Linolenic   | trace              | 164                            | 8.4          | 172   |

<sup>\*</sup> mg/100 g fresh tea leaves, sample in June, 1975.

Table 2. Changes in linoleic and linolenic acid in tea leaves during blending

| Fraction        | Linoleic acid |       | Linolenic acid |       |
|-----------------|---------------|-------|----------------|-------|
|                 | 0*            | 3     | 0              | 3     |
| Free fatty      | trace         | trace | trace          | trace |
| acid            | trace         | trace | trace          | trace |
| Non-ionic lipid | 55.6†         | 23.5  | 218.3          | 99.3  |
|                 | 18.5‡         | 15.8  | 164.0          | 145.8 |
| Phospholipid    | 16.8          | 13.3  | 11.5           | 10.5  |
|                 | 13.8          | 3.5   | 8.4            | 1.4   |
| Total lipid     | 72.4          | 36.8  | 229.8          | 109.8 |
|                 | 32.4          | 19.3  | 172.4          | 147.2 |

<sup>\*</sup>Blending time (min). Zero blending time: fractions prepared from leaves inactivated by heating at 80° for 15 min. † Sampled on the 6th June. ‡ Sampled on the 26th November.

40° for 10 min. 6 ml of headspace volatile thus obtained was analyzed by GLC under the above conditions.

Preparation of fatty acids. Crude lipid was extracted with CHCl<sub>3</sub>-MeOH (2:1) 1.1. from blended tea leaves (200 g). Fatty acids from each lipid fraction were prepared according to the method of ref. [11] and were converted to Me esters with CH<sub>2</sub>N<sub>2</sub>-Et<sub>2</sub>O. Samples of the esters were analyzed by GLC with 20% PEG-adipate on Chromosorb W, column temp, 180°.

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